no evidence, however, that this is true; theoretically, field-collected colonies could become laboratory adapted within their first few generations on oleander scale in the insectary. If commercial insectaries wish to maintain the R strain's purity, they should not add field-collected parasites to the R colony, unless those parasites are resistant to Sevin (see below).

Conclusions

The laboratory-selected R strain of A. melinus is ready for practical use, and one commercial insectary has already received the strain for mass rearing and release. If adequate resistance can be maintained in the \bar{R} strain in an insectary throughout the spring and summer release periods, then orchards that receive the parasites may acquire a more permanent population of Sevin-resistant parasites. If resistant populations predominate in release areas, commercial insectaries could collect A. melinus from these areas to replenish their colonies yearly, and periodic selection of these parasites could maintain Sevin resistance in the insectary.

The R strain could provide better control of California red scale than susceptible parasites in pesticide-treated citrus orchards and could prevent pest resurgences or secondary pest outbreaks. The results: (1) fewer applications, or lower rates of application, of pesticides to control red scale; (2) lower control costs to growers, and (3) a reduced pesticide load in the environment.

Commercial insectaries and growers interested in obtaining the Sevin-resistant strain can contact Foothill Agricultural Research, 510¹/₂ West Chase Drive, Corona, CA 91720; phone: (714) 371-0120.

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Site location for bermudagrass and redroot pigweed control in Humboldt County, 1989. The picture was taken 10 weeks after solarization began. The bare strips are where tarps were removed after 6 weeks; tarps still in place indicate the 14-week solarization.

Perennial weeds respond to control by soil solarization

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In soil solarization (soil tarping), solar energy is used to reduce soil pests. Clear polyethylene tarps may be used singly to cover pre-formed beds or they can be glued together to cover broad areas. Weed control is most effective when prepared beds are irrigated before being covered with polyethylene; however, when beds have drip lines, irrigation can take place after polyethylene is installed. Moisture under the tarp helps conduct heat and stimulates weed seeds to germinate; the heat then kills the seeds.

Winter annual species, such as annual bluegrass, common sowthistle, and shepherdspurse, are readily controlled by solarization; winter annual grasses are especially sensitive. Many summer annual species, including barnyardgrass and pigweed, are also controlled; the exception, sweet clover, is only partially controlled.

Perennial weeds are more difficult to control than annual weeds because their

Soil solarization in summers in Central Valley and near-coastal sites controlled bermudagrass and johnsongrass covered by polyethylene tarps. Field bindweed was also controlled during solarization, but regrowth occurred. To maximize control and reduce the "edge" effect – the regrowth of weeds at the edges of fields when they are not covered by tarps – the soil must be completely covered with intact polyethylene.





TABLE 1. Preparation for weed control with polyethylene application (and removal) at five California sites

	Solarization site						
	Bermudagrass		Johnsongrass		Field bindweed		
	1	2	3	4	5		
Treatment before tarp installation	Turf mowed or rototilled	Oat stubble/ bermuda mowed, raked, rototilled	Rototilled	Disked, listed, bedded, mechanically rototilled	Disked, listed, bedded, mechanically rototilled		
Irrigation	Sprinkler	Hand-watered with hose	Sprinkler plus hand-watered with hose	Sprinkler	Sprinkler		
Tarp applied	August 8	June 15	July 3	July 7	July 6		
Tarp removed (weeks after application)	6	6 or 14	9 or 12	6 or 8	6 or 9		
Avg. maximum air temp.	86.4°F (30.2°C)	6 wks, 78.6°F (25.9°C)	9 wks, 87.7°F (30.9°C)	6 wks, 97.7°F (36.5°C)	6 wks, 97.7°F (36.5°C)		
Avg. solar radiation (langley)	574	NA*	NA	676.6	676.6		

Below left, johnsongress control after 6 weeks of solarization in Davis, California. Bare strips are where beds were solarized and the tarps removed.

underground vegetative structures — rhizomes, tubers, or bulbs — allow them to survive most nonchemical controls, including cutting, cultivation, and mowing. Use of solarization, therefore, could be a desirable alternative to chemical control.

Bermudagrass, a perennial grass, forms rhizomes (underground stems) capable of covering a large area and forming numerous new plants when cut into pieces. In nontillage culture — as in orchards or landscapes — bermudagrass rhizomes may extend 6 inches deep or more. In cultivated areas, rhizomes are usually found to the depth of cultivation. These rhizomes, as well as surface stems (stolons), must be killed to eliminate this species.

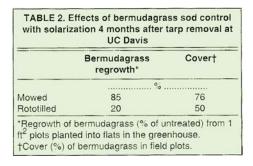
Johnsongrass, another perennial grass, also forms rhizomes. Like bermudagrass rhizomes, johnsongrass rhizomes usually grow to plow depth in cultivated, lighttextured soils. In nontilled areas, rhizomes often grow 4 to 6 inches deep.

Field bindweed, a broadleaf perennial weed, has a storied rootstock system that can extend as deep as 8 to 12 feet in the soil; rootstocks, however, are predominantly found in the top 18 inches of the soil profile.

The following soil solarization studies evaluated the control of the perennial weeds bermudagrass, johnsongrass, and field bindweed. Information on the preparation for all sites is summarized in table 1.

Bermudagrass studies

Control of bermudagrass was evaluated at three sites: at the University of California at Davis (site 1) and at two sites 22 miles east from the coast in Humboldt County's Eel River Valley (sites 2 and 3). The UC



Davis experiments took place in 1980, while Humboldt County experiments took place in 1989 and 1990.

The Davis location had established turfgrass sod before start of the study in 1980. Some beds at the site were mowed to 1/2 inch and others were rototilled with a garden rotovator to 4 inches deep before the tarps were laid to test whether there would be a difference in control results between mowing and rototilling. The site was sprinkler-irrigated before tilling and irrigation ceased during the solarization period. A slight decline in the slope (7%) to the north reduced the sun angle. Polyethylene tarps (2 mil) were sealed at the edges August 8, 1980, by removing soil to make a trench, inserting the tarps into the trench, and covering the edges. The tarps, pulled tightly to keep them as close to the soil as possible, were then left undisturbed for 6 weeks.

The plots, 5 feet by 5 feet, were replicated four times in a randomized complete block design. After the tarps were removed, soil samples containing bermudagrass rhizomes and other plant parts were sampled from a 1-square-foot area to 6 inches deep in each plot and placed on flats in the greenhouse; bermudagrass regrowth was then evaluated. Regrowth of bermudagrass in the field was visually evaluated as a percentage cover of the plot and compared with untreated turfgrass adjacent to the outside edge of each plot.

Site 2 in Humboldt County was heavily infested with bermudagrass and had been cropped with oats before the study began. The plots were mowed, raked free of forage, and rototilled 4 inches deep June 15, 1989. Before tarping, the soil surface was watered by hand (with approximately 1 inch of water). There was no moisture below 2 to 4 inches of the soil at the time of treatment and no subsequent application of water. Polyethylene tarps (2 or 1.5 mil) were hand placed on the plots June 15, 1989. Plots were 5 feet by 10 feet and replicated four times using a randomized complete block design. The tarps were sealed on the edges as described in the first study. Use of a standard single layer of polyethylene (1.5 mil thick) covered by a second layer was evaluated. The tarps remained on the soil for 6 or 14 weeks before removal. On September 29, 1989, bermudagrass control was evaluated both by estimates of percent cover and by visual evaluation of live rhizomes excavated from the upper 1 foot of the plots (white, turgid rhizomes = alive; brown rhizomes = dead) (table 1). Control of redroot pigweed was also evaluated visually.

In 1990, another study (site 3) was established adjacent to the 1989 Humboldt County site. The field was disked and TABLE 3. Bermudagrass and redroot pigweed control with soil solarization for 6 and 14 weeks and with one and two layers of polyethylene in Humboldt County, 1989

	Number		Polyethylene thickness	Weed control		
	of tarp	Solarization		Bermudagrass		Redroot
Treatment	layers	duration		Control*	% Cover [†]	pigweed
		wks	mil			
Solarization	single	6	2§	9 b	2b	10b
Solarization	double	6	2	8 b	15b	· 10b
Solarization	single	14	2	9 b	4b	10b
Solarization	double	14	2	10 b	0.8b	10b
Solarization	single	14	1.5¶	9 b	7b	10b
Untreated	-0-	-0-	-0-	3 a	65a	5a

*Visual evaluation without knowing treatments, of shoot growth and white, turgid, rhizomes, mean of 4 replications. 1 = no control; 10 = complete control.

†Estimated cover of the plot (excluding regrowth from outside edges).

‡Visual rating. 1 = no control; 10 = complete control.

§Ultraviolet stabilized polyethylene, Monsanto Agricultural Products.

 $\P U traviolet stabilized polyethylene; Visqueen <math display="inline">{}^{\textcircled{R}}$, Ethyl Corporation.

Note: Means in each column followed by the same letter do not differ significantly at the 5% level (Duncan's multiple range test).

TABLE 4. Bermudagrass control with soil solarization for 9 or 12 weeks in Humboldt County, 1990

Treatment	Solarization duration	Tarp layers	Bermudagrass cover*		Bermudagrass control
			Whole plot	Inside a 6-in margin	Rhizome dry weight†
	wks	no.	%		g
Solarization	9	1	8	1b	0.0b
Solarization	12	1	7	Ob	0.0b
Solarization	9	2	2	Ob	0.0b
Untreated	0	0	100	98a	12.3a

*Visual evaluation September 27, 1990, when the 12-week tarp was removed. Mean of three replications. †Rhizomes removed from an area 1 foot wide, 1 foot deep, and 3 feet long, air dried, and weighed.

Note: Means in each column followed by the same letter do not differ significantly at the 5% level (Duncan's multiple range test).

TABLE 5. Established johnsongrass control with soil solarization for 6 or 8 weeks in Davis, California, 1990

Treatment		Johnsongrass control			
	Solarization duration	Johnsongrass cover	Johnsongrass shoots†	Johnsongrass rhizome‡ dry wt	
	wks	%*		g	
Solarization	6	Ob	1b	5b	
Solarization	8	Ob	Ob	Ob	
Untreated	0	58a	46a	615a	

*Visual evaluation of % cover on bed top (October 18, 1990).

*Number of johnsongrass shoots per 1 square meter (three subsamples per plot) evaluated May 8, 1991. *Rhizomes removed from area, 1 foot wide, 1 foot deep, and 3 feet long, air dried August 15, 1991.

Note: Means in each column followed by the same letter do not differ significantly at the 5% level (Duncan's multiple range test).

TABLE 6. Field bindweed control with soil solarization for 6 or 9 weeks in Davis, California, 1990

Treatment		Field bindweed control			
	Solarization duration	Fresh weight* 10/1	Seedlings† 10/10	Top growth control‡ 10/18	
	wks	g			
Solarization	6	27.0b	0.6b	7.8a	
Solarization	9	0.0b	0.8b	7.8a	
Trifluralin, 0.75 lb/ac	—	429.2a	0.3b	7.2a	
Untreated	0	543.5 a	4.2a	1.0b	

Grams of field bindweed harvested from 30 feet of 60-inch bed.

[†]Mean number of seedlings per 1 ft² quadrat.

‡Visual evaluation of field bindweed top growth: 1 = no control; 10 = complete control.

Note: Means in each column followed by the same letter do not differ significantly at the 5% level (Duncan's multiple range test).

sprinkler-irrigated (with approximately 4 inches of water) before tarps were laid July 3, 1990. The plots were watered by hand to field capacity just before the tarps were sealed. As in the neighboring experiment, single or double layers of 1.5-mil polyethylene were laid. The tarps remained on the soil for 9 or 12 weeks. Plots were 5 feet by 8 feet and replicated three times. Bermudagrass control was visually evaluated in the entire plot. Live rhizomes from an area in each plot 1 foot wide, 3 feet long, and 1 foot deep were removed, air dried, and weighed.

Johnsongrass study

Controlling johnsongrass with solarization was evaluated in a heavily infested field on the UC Davis farm (site 4). After the soil was disked and listed, beds were formed and sprinkler-irrigated with 4 inches of water. Following irrigation, the beds were mechanically rototilled and shaped into finished 5-foot-wide beds before tarps were laid. Plot size was 5 feet by 35 feet and replicated four times in a randomized complete block design. On June 20, 1990, the tarps were applied to moist soil with a tractor-mounted mulching machine; they remained on the beds for 6 weeks in one treatment, and 8 in another, before they were removed. Johnsongrass shoots were counted in three subsample square meter quadrants on May 8, 1991 and the mean used for analysis. Control of johnsongrass rhizomes was determined by visual evaluation of regrowth in the plots and by measuring the dry weight of rhizomes collected from an area 1 foot wide, 3 feet long and 1 foot deep in each plot.

Field bindweed evaluation

A site at UC Davis (site 5) was heavily infested with established field bindweed. Beds were prepared like those in the johnsongrass site. The 1.5-mil polyethylene was applied to pre-formed 5-foot beds July 7, 1990 with a mechanical layer. The tarps remained on the beds for 6 weeks in one treatment and 9 weeks in another. Trifluralin at 0.75 pound active ingredient per acre (a.i./ac) was applied as a comparison treatment to the bed surface and sprinklerincorporated with 1 inch of water. Field bindweed growth from established plants was harvested from each plot October 1, 1990 and weighed. Field bindweed seedlings were counted 34 days after the tarps were removed and coverage of established shoots was visually evaluated 42 days after their removal.

Solarization's effects

Solarization reduced regrowth and coverage by rototilled or closely mowed, trash-free bermudagrass sod (table 2) at site 1 (UC Davis). Greater control of bermudagrass was achieved when the sod was rototilled than when it was mowed. Bermudagrass was effectively controlled with a single sheet of 1.5- or 2-mil polyethylene tarp when applied in summer at both sites in Humboldt County; a double layer was found to be no more effective than a single layer (tables 3 and 4). Part of the decrease in control at the Davis site compared with the cultivated Humboldt County sites was due to reduced radiation on the north-facing slope. Where holes developed in the polyethylene, bermudagrass survived and grew. However, rhizomes were killed where the polyethylene remained intact or was patched with clear plastic tape. Bermudagrass regrew in treated plots from the edges (particularly apparent on the north edge of the plot) where the soil covered the polyethylene (table 4). Rhizome regrowth occurred from rototilled or mowed plots but to a lesser extent in rototilled soil greenhouse tests (table 2).

Johnsongrass was effectively controlled with 2-mil polyethylene tarps for 6 and 8weeks at Davis (table 5). Mature rhizomes and seedlings were controlled to 8 inches deep, the maximum depth rhizomes were found. Like bermudagrass, johnsongrass regrowth occurred if a large clump of rhizomes was not completely covered with polyethylene. Thus, there is a distinct "edge" effect when tarping to control both perennial grasses.

Controlling field bindweed with soil solarization was not as effective as perennial grass control. The number of field bindweed seedlings was reduced with solarization, but the seedlings were not eradicated (table 6). Regrowth from established field bindweed was suppressed for 6 weeks after solarization. Top growth was visually evaluated 17 days after tarp removal and was not different between tarping treatments but was significantly reduced from untreated plots. Trifluralin at 0.75 lb a.i./ac, applied to the soil surface, did not suppress field bindweed fresh weight 12 weeks after application. After top growth was removed, seedling numbers and new top growth were suppressed.

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